# CosmoPAGE Bis-tris Precast Gel
## *- Bis-tris gel, 15 wells –*

**12 %, Cat. No. NU001215**
**4-12 %, Cat. No. NU041215**

**Introduction**
CosmoPAGE Bis-Tris Precast Gel is a high-performance and easy to use precast polyacrylamide gel for electrophoresis in Bis-Tris buffer system (MOPS or MES). The optimized gel formula allows CosmoPAGE Bis-Tris Precast Gel to show improved resolution, accurate results, and an extended shelf-life over conventional gels. CosmoPAGE Bis-Tris Precast Gels are available in gradient (4 to 12%) and fixed (12%) concentrations of polyacrylamide in 15-well formats.

**Product Information**

| Product | Cat. No. | Gel % / # of well | QTY |
|---|---|---|---|
| CosmoPAGE Bis-tris Precast Gel | NU001215 | 12% / 15 wells | 10 gels |
| CosmoPAGE Bis-tris Precast Gel | NU041215 | 4 - 12 % / 15 wells | 10 gels |

**Storage**
4 °C

**Shelf Life**
12 months

**Features**
- Compatible with Bio-Rad Mini-PROTEAN® CORE
- No combs and tape
- Sharp, straight bands
- Numbered well; loading volume up to 28 µl/well

**Procedures**

**Running Buffer**
Dissolve CosmoPAGE MOPS Running Buffer in 1000 ml of deionized water to make the 1X MOPS running buffer. Or prepare 10X stock running buffer with the following recipe:
10X MOPS running buffer:
Tris base 60.6 g, MOPS 104.6 g, SDS 10.0 g, EDTA 3.0 g.
Deionized water to 1000 ml.
10X MES running buffer preparation:
Tris base 60.6 g, MES 97.6 g, SDS 10.0 g, EDTA 3.0 g.
Deionized water to 1000 ml
*Always use fresh 1X running buffer

**Sample Preparation**
Mix sample with 2X sample buffer. Heat the diluted samples at 90-95°C for 5 min.
Recipe of 2X sample buffer with reducing agents:
62.5 mM Tris-HCl pH 6.8, 2% SDS, 25% (v/v) glycerol, 0.01% bromophenol blue, and 100 mM DTT (or 5% β-mercaptoethanol) as reducing agent.

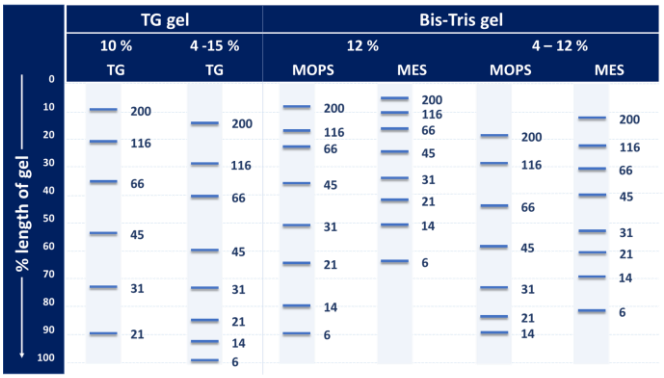
**Setting CosmoPAGE Precast Gel into Bio-Rad Mini-PROTEAN® Core**
Totally the same way to place CosmoPAGE precast gels and Bio-Rad Mini-PROTEAN gels; no adaptors required.

**Running Conditions**

| | 100 V | 150 V | 200 V |
|---|---|---|---|
| Running time | 90 min. | 60 min. | 35 min. |
| Current | | | |
| Initial per gel | 15-20 mA | 50-60 mA | 100-110 mA |
| Final per gel | 5-10 mA | 20-30 mA | 30-40 mA |
| TEMP. | 25°C | 25-35°C | 25-35°C |

Running time might vary depending on power supply and the temperature of gel, running buffer and lab.

**Gel Migration chart**




Migration patterns of Unstained Protein Markers (#29458-24)

**Gel staining**
1. Insert a cassette opener into four corners of cassettes.
2. Carefully **pry but not twist** the opener to separate two plates.
3. Gently remove the gel from the plate.
4. Follow a procedure of most popular staining methods, such as Coomassie dyes (R-250 or G-250), Silver-stain solution, and fluorescent.

**Gel blotting**
Follow the general guidelines for your blotting apparatus; compatible to wet, semi-dry, or dry transferred to PVDF or nitrocellulose membrane.

**Technical Support**
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